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All Animal Care Personnel  
Veterinarians, All Animal Program Personnel  
To Establish the Proper Guidelines for the Quarantine of Rodents

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1. To prevent the introduction of rodent pathogens including environmental opportunistic agents into established rodent colonies.
  
1. The veterinarians oversee all aspects of animal health, and are assisted by all program staff.
2. The Assistant Directors are responsible for ensuring that Facility Managers implement all practices.
3. The Facility Manager is responsible for ensuring that all technical and animal care staff are adequately trained and experienced in rodent quarantine procedures.
  
1. Rodents from the \_\_\_\_\_ . The Fiscal & Business Specialist is responsible for ensuring that current health statuses of the approved vendor colonies from which requested animals originate are SPF/VAF for infectious agents on the exclusion list. All animals should be acclimated a minimum of seven days prior to use.
2. Rodents from non-approved sources including other commercial vendors and from academic institutions, \_\_\_\_\_ as described below.
3. Access to quarantine rooms/facilities is limited to \_\_\_\_\_. Research personnel are not allowed in quarantine areas unless escorted by a veterinarian or designee.
4. \_\_\_\_\_ ensuring the procedures outlined in the quarantine schedule, performance of health evaluations, interpretation of test results, and when all is successfully completed, the released of animals from quarantine. Technicians are assigned by the facility manager or designated supervisor and assisted by other assigned animal care staff at each facility in the collection of samples.
5. Just Dedicated quarantine scrubs and shoes, disposable gown, shoe-covers, bouffant cap, and gloves, or Tyvek® coverall, bouffant cap if Tyvek® hood is not worn, and gloves are



- a. Remove microisolator top, empty feed from wire rack and the contents of the water bottle onto the cage bedding, and either replace the uncapped water bottle in the microisolator bottom or cap the water bottle and replace in the wire rack.
- b. ®  
so that it can be  
readily observed and replace the microisolator top. Do not clamp IVC microisolator tops.
- c.

3. Fur mite sample collection procedures can be found at:  
<https://www.idexxbioresearch.eu/new-page-3>

results are positive the veterinarian/facility m



3. The relocation of mice to or from the CPH is accomplished in autoclaved commercial filtered shipping boxes that are exterior surface decontaminated with Oxivir Tb® upon arrival at the remote area (e.g., microscopy core). Each transport box will contain a single “home” microisolator cage of mice.
4. Shared equipment in common use areas are either surface decontaminated with Oxivir Tb® (e.g., multiphoton microscope stage, inhalational anesthesia nose cone) for the appropriate contact time before and after each transport box (i.e., “home” microisolator cage) of mice are used (e.g., imaged) in accordance with .
5. Whenever SPF/VAF mice require relocation to other facilities for use of resources unique to that facility (e.g., MRI image acquisition), a **Request to Relocate &/or Reassign Research Animals** must be submitted. Scheduling of mouse use is made when other mice are not present in the immediate area, and surfaces and common equipment are decontaminated before and after use in accordance with .