

STANDARD OPERATING PROCEDURES
DIVISION OF COMPARATIVE MEDICINE
UNIVERSITY OF SOUTH FLORIDA

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TITLE:	Common Procedural Area Decontamination and Room Duties
SCOPE:	All Animal Program Personnel
RESPONSIBILITY:	Facility Manager, Animal Program and Research Personnel
PURPOSE:	To Outline the Proper Maintenance of Common Areas in Animal Facilities.

I. PURPOSE

1. To describe decontamination procedures to prevent the spread of microbial agents that may cause sub-clinical and clinical diseases that could jeopardize the validity and reproducibility of research data, or complicate its interpretation, or cause zoonotic concerns.

II. RESPONSIBILITY

1. The Facility Manager or designee ensures that common areas, corridors, animal procedural areas, core facilities, and equipment located in their facility are cleaned regularly, disinfected appropriately, the efficacy of sanitation documented, and available to research personnel as requested.
2. It is the responsibility of the individual(s) using a common procedural area or core facility to disinfect all equipment and work surfaces that may come in contact with animals prior to and after each use as described below.
3. The Facility Manager or designee assigns personnel and intervals of sanitation to maintain common areas, corridors, animal procedural, and core facilities areas as appropriate for the area/equipment and frequency of use.
4. The Facility Manager or designee ensures implementation of all procedures.

III. GENERAL DECONTAMINATION PROCEDURES

1. All portable and fixed equipment as well as surfaces that come in contact with animals must be decontaminated prior to and after each episode of use by the personnel using

a. Decon- Spore 200 Plus-

Active ingredient - peroxyacetic and hydrogen peroxide solution
Preparation - 6.0 ml/gal water
Safety - strong oxidizer and corrosive-use in a well ventilated area
PPE - gloves and eye protection when handling/mixing concentrate
Uses - sanitizing floors throughout the animal facility
Specific uses- Mopping floors and floor machine
Efficacy - Staphylococcus aureus, Enterobacter aerogenes, Escherichia coli, Listeria monocytogenes, Salmonella typhimurium, Psuedomonus aeruginosa, and Saccharomyces cerevisiae
Contact time - 5 minutes at ambient temperature.

b. Oxivir

Active ingredient -accelerated hydrogen peroxide solution
Preparation - Oxivir Tb and Oxivir wipes are ready to use. Oxivir Five 16 concentrate is diluted 1 part concentrate: 16 parts water
Safety - excellent safety, avoid contact with eyes and skin
PPE - gloves and eye protection when handling/mixing concentrate
Uses -general disinfectant for hard non-porous surfaces in laboratory animal procedural areas and for surfaces that come in contact with animals.
Specific uses- forceps or gloved hand-dipping between handling animals, biosafety cabinet chambers, animal restrainers/containers used in imaging (e.g., irradiator pie cages, anesthetic chambers/nose cones, MRI/Xenogen chambers, behavioral equipment)
Efficacy - virucide, bactericide, fungicide, mildewcide.
Contact time - Disinfects in 1 minute; Tuberculocidal in 5 minutes

NOTE- Personnel handling disinfectants should be familiar with product directions and MSDS

4. Animal care staff is assigned to specific common procedural and core facility areas and are responsible for ensuring the cleaning, disinfection, and documentation of efficacy of sanitation of their assigned areas at intervals specified by this procedure. This procedure describes minimum requirements, additional duties and frequencies are at the Facility Manager 's discretion.
5. Floors are swept and sanitized with Decon- Spore 200 Plus and trash is emptied.
6. Surfaces that come in contact with animals are disinfected with Oxivir Tb.
7. Surfaces unlikely to come in contact with animals (e.g., counter tops, HVAC ducts, fixed equipment, carts, walls, and fixtures are wiped clean using Oxivir Tb.
8. Air filters, when present, are changed as required by monthly checks to ensure a dust and debris-free environment.
9. Mop heads and buckets are rinsed after each use and mop heads are cleaned/sanitized in the rack washer at least weekly . At the CAMLS, mop-heads are hand washed, rinsed, and sanitized by hand.

IV. EQUIPMENT DECONTAMINATION PROCEDURES

1. Xenogen-
 - a. Imaging chamber floor is sanitized with Oxivir Tb wipes and paper liners that come in contact with animal(s) are changed before and after each cage of animals is imaged.
 - b. Anesthesia chamber is sanitized with Oxivir Tb before and after each cage of animals is anesthetized.

2. X-ray irradiator -
 - a. Chamber floor is wiped clean and sanitized with Oxivir wipes and absorbent liners that come in contact with animal(s) are changed before and after each cage of animals are exposed.

3. MRI-
 - a. Animal holder/restrainter is cleaned and sanitized with Oxivir wipes and after each cage of animals is imaged.
 - b. Anesthesia chamber is sanitized with Oxivir Tb before and after each cage of animals is anesthetized.
 - c.

- d. Vaporizers checked twice daily and filled as needed
- e. Additional O₂ tanks available in the room
- f. Scavenging canisters weighed at end of day

9. Behavioral Apparatus-

- a. Behavioral apparatus are sanitized with Oxivir Tb between cohorts of animals from different investigators and after use each day.
- b. Behavioral apparatus are also disinfected with Oxivir wipes or alcohol spray or wipes, preferably after each animal is tested (since the scent of another animal may complicate the interpretation of data), but at least after every cage of animals tested.

10. Laser Doppler -

- a. Counter is sanitized with Oxivir wipes and black paper liners that come in contact with animal(s) are changed before and after each cage of animals is imaged.

11. Olympus IV 100 Scanning Microscope -

- a. Stage is wiped clean and sanitized with Oxivir wipes and paper liners that come in contact with animal(s) are changed before and after each cage of animals are exposed.
- b. Micro Probe lenses that come in contact with animals should be cleaned/ disinfected after each animal with alcohol only. Objective lenses that happen to come in contact with animals should also be disinfected after each animal with alcohol only.
- c. Anesthesia nose cone is sanitized with Oxivir Tb before and after each cage of animals is anesthetized.

12. CODA non- invasive blood pressure system -

- a. Occlusion cuffs can be gently rinsed with water and cleaned with germicidal or antiseptic soap and water before and after each cage of animals. Do not use solvents or alcohol to clean cuffs.
- b. Since cuffs cannot be adequately sanitized, they will be dispensed for use on an investigator's own animals to minimize the spread of infectious agents throughout a facility.

13. Confocal and Multiphoton Microscopes -

- a. Stage is wiped clean and sanitized with Oxivir wipes before and after each cage of animals are exposed.
- b. Anesthesia nose cone is sanitized with Oxivir Tb before and after each cage of animals is anesthetized.

14. PET/SPECT/CT

- a. Animal b

